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Estimating neural signal dynamics in the human brain

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Although brain imaging methods are highly effective for localizing the effects of neural activation throughout the human brain in terms of the blood oxygenation level dependent (BOLD) response, there is currently no way to estimate the underlying neural signal dynamics in generating the BOLD response in each local activation region (except for processes slower than the BOLD time course). Knowledge of the neural signal is critical if spatial mapping is to progress to the analysis of dynamic information flow through the cortical networks as the brain performs its tasks. We introduce an analytic approach that provides a new level of conceptualization and specificity in the study of brain processing by non-invasive methods. This technique allows us to use brain imaging methods to determine the dynamics of local neural population responses to their native temporal resolution throughout the human brain, with relatively narrow confidence intervals on many response properties. The ability to characterize local neural dynamics in the human brain represents a significant enhancement of brain imaging capabilities, with potential applications ranging from general cognitive studies to assessment of neuropathologies.

Keywords: fMRI, cortical processing, human, visual, neural dynamics, blood oxygen level dependent

INTRODUCTION

The basic capability of functional magnetic resonance imaging (fMRI) is mapping the spatial organization of the cortical and subcortical responses throughout the brain to a variety of stimulus and behavioral paradigms. The blood oxygen level dependent (BOLD) signal measured by fMRI is often considered to have high spatial resolution (up to a quarter of a million voxels), but low temporal resolution (0.5–5 s) relative to other methods for mapping human brain function (such as high-density EEG analysis). Of course, even the BOLD spatial resolution is low relative to single-unit neurophysiological recoding, but this is largely attributable to the signal/noise ratio. In-plane spatial resolution for functional imaging as high as 150 μm has been reported at 9.4 T in cat cortex (Harel et al., 2006) and less than 1 mm $\times$ 1 mm $\times$ 1 mm voxels have been achieved in human (Barth and Norris, 2007). As improved technologies come on line, such high spatial resolution is expected to be continually increased, as exemplified by the “inverse imaging” approach of Lin et al. (2008).

The temporal limit is not so tractable. The inherent time constants of the BOLD signal are about 5 s, several orders of magnitude slower than the neural signal that is driving it, even for the most recent fast acquisition protocols (Lin et al., 2008; Grotz et al., 2009). For a reasonable recording time and voxel size (e.g., 3 mm $\times$ 3 mm $\times$ 3 mm) the signal/noise ratio even for the most effective stimuli rarely exceeds 20:1. At such signal/noise ratios, deconvolution of the BOLD response can improve its temporal resolution by about a factor of 5 (Glover, 1999; Logothetis, 2003). However, that leaves the temporal resolution for the underlying neural signal at about 1 s, which is far short of what is required to measure typical neural delays. Estimation of single-parameters of the waveform, such as response delay alone, can improve the temporal resolution for the neural signal delays to 100 ms or better for narrowly targeted brain regions (Menon et al., 1998; Henson et al., 2002) but this requires the assumption that the BOLD signal

has a unitary waveform, which is often not the case (Aguirre et al., 1998; d’Avossa et al., 2003; Handwerker et al., 2004; Likova and Tyler, 2007). Even minor deviations from a stable waveform violate the assumptions of such single-parameter analysis and invalidate the delay measure.

Indeed, most commonly used fMRI analysis techniques, such as the SPM analysis package, employ the convolution approach. Convolution is based on the assumption of a unitary BOLD waveform kernel that generates the straightforward prediction of the BOLD response waveform for any stimulus type or duration in any brain area. In fact, however, major deviations from a standard BOLD waveform may be found, even in the same cortical regions, for variations in stimulus conditions. d’Avossa et al. (2003), for example, reported strong differences in waveform when the response to the motion or color of a cue/stimulus pairing was modulated by attention. Such local waveform differences must derive from differences in the neural signals driving the BOLD activation, since the metabolic and hemodynamic processes that mediate the paramagnetic signals should be invariant within a given cortical region.

To study the neural involvement in generating differential BOLD response waveforms within the same cortical regions, Likova and Tyler (2007) developed an “instantaneous stimulus paradigm” to evoke BOLD responses to instantaneous stimulus transitions. It is typically assumed that, despite their perceptual differences, such “instantaneous” stimuli would all generate the same BOLD waveform (effectively equivalent to the hemodynamic response function, HRF) for all different stimulus types. Thus, revealing any significant deviation from that prediction in the BOLD waveform elicited in one and the same cortical area can contribute to revealing the specifics of the underlying neural processing and enhance the understanding of the recurrent network of extended perceptual responses to complex stimulus configurations. Indeed, the instantaneous stimulus paradigm generated a wide variety of BOLD waveforms. Moreover, the data show dramatic differences in the BOLD waveforms properties

(e.g., latency, sign, amplitude, and width) even *within the same* brain areas as a function of the stimulus type. As there can be no vascular heterogeneity within the same area, these dramatic waveform variations must be attributed to differences in the underlying *neural dynamics*, not to spatial variations in the HRF. These results further imply that fMRI signals contain much more information about the neural processing than is commonly appreciated, and thus have the potential to capture them through an appropriate approach.

However, there is at present no method of transcending the BOLD temporal limitations in order to estimate the dynamics of the neural signals underlying the measured fMRI waveforms. The goal of the temporal analysis we propose is therefore to provide a method for the time-resolved estimation of the neural signals underlying the particular characteristics of the temporal BOLD waveforms for a particular stimulus processed by a particular cortical region. The philosophy of this approach is to utilize the information available from neurophysiological studies of the neural population dynamics and biochemical studies of the metabolic pathway coupling to the measurable blood response to provide Bayesian priors as to the likely temporal structure of the component neural signals. We propose a *non-linear dynamic forward optimization* (NDFO) approach to provide a compact account of the measured waveform with the minimal number of neural predictors, based on prior knowledge of the expected temporal properties of neural signals and of their consequent metabolic demand. This approach is reminiscent of the spectral analysis of the composition of stars, in which each chemical element has a characteristic pattern of emission lines (the Bayesian priors) and the net spectrum is the sum of an unknown mixture of these predetermined patterns. The result is the ability to specify the prevalence of each element in the otherwise inaccessible star. In the case of the neural signals, the goal is to estimate the amplitude and time course of each of the neural components whose metabolic

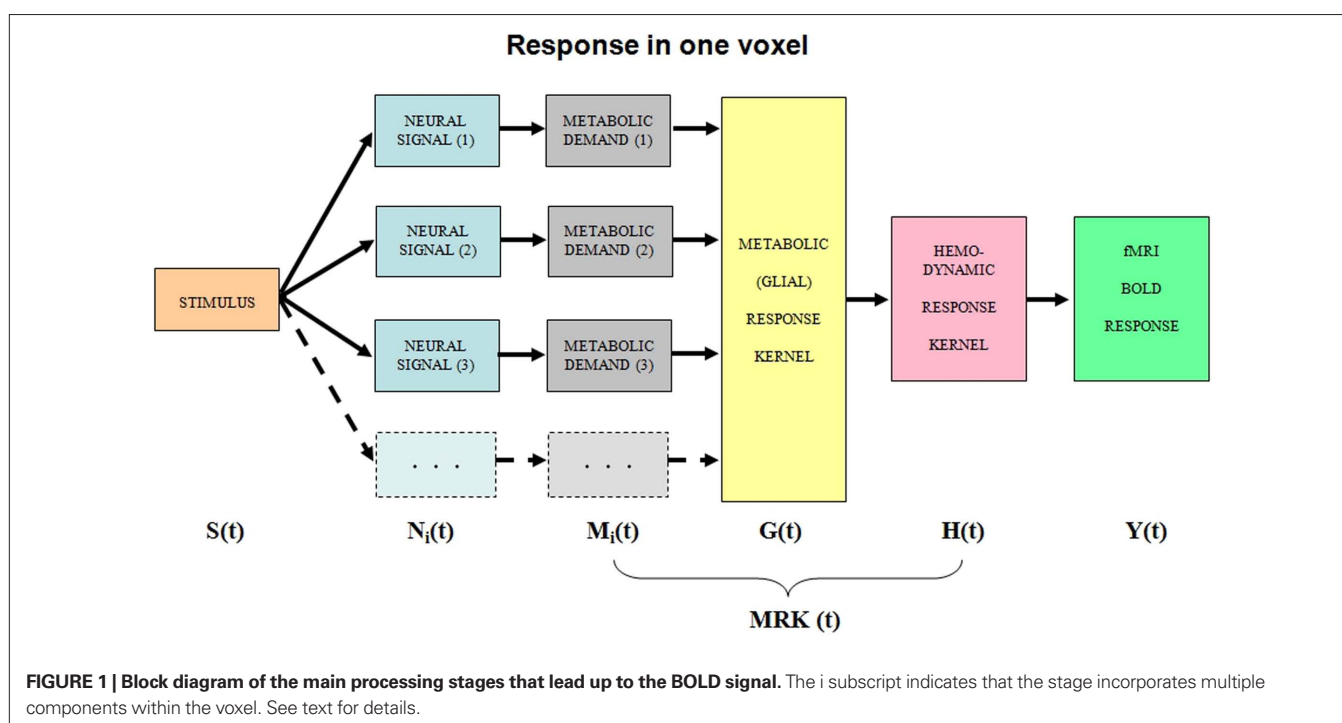
effects, when summed, account for the measured BOLD waveform for a particular stimulation condition and cortical region. Here the predictors are non-linear because there is a non-linear relationship between the neural responses and the metabolic demand that they generate, but the summative property of the paramagnetic signals throughout a voxel implies that we can assume that the component metabolic demands sum linearly together.

THEORETICAL OVERVIEW

To start the analysis, we may develop a specific model structure of the processes leading to the BOLD paramagnetic signal of fMRI recordings. We note that this model is focused on the non-linearities that are likely to affect the process of estimating neural responses from BOLD waveform properties, but simplifies aspects of the biophysical processes that will not be resolvable by the NDFO technique. As such, it is substantially elaborated relative to the linear convolution analyses of Friston (1997) and Friston et al. (1998, 2000), but somewhat condensed in comparison with the recent biophysical/metabolic derivations of Mechelli et al. (2001), Buxton et al. (2004), or Sotero and Trujillo-Barreto (2007, 2008).

We will treat the *neural responses* within each voxel as generated by sets of homogeneous populations with similar signal waveforms $N_i(t)$ within each population (**Figure 1**). Each neural response then generates a *local metabolic demand* $M_i(t)$ that may have a non-linear relationship to the neural signal waveform (Chatton et al., 2003).

The metabolic demand has been shown to have a short time course, on the order of the membrane response to a single pre-synaptic spike (Magistretti and Pellerin, 1999; Magistretti, 2009). The *integrated metabolic demands* are met primarily by the astrocytes, which integrate the required energy consumption over time and space and make a *complementary oxygen demand* $G(t)$ on the adjacent vasculature. The *hemodynamic processes* $H(t)$ provide



the requisite oxygen exchange to replenish the energy depletion in the astrocytes and other intermediary processes. The fMRI BOLD analysis provides an estimate $Y(t)$ of the ratio of oxygenated to deoxygenated hemoglobin in the blood complement of a given voxel. The *post-neural* processing stages are often modeled as a *linear hemodynamic response kernel* convolved with the presumed neural signal. However, this approach overlooks the key role of the *pre-hemodynamic* processes of the *glial* and other intermediaries. To incorporate the contributions of these intermediary processes, we will term it a “*metabolic response kernel*” (MRK¹) incorporating *both* the glial and hemodynamic components of the metabolic recovery processes. The MRK will be convolved with a non-linear transform of the presumed neural signal to provide an estimate of the neural metabolic demand that is being met by the combined glial and hemodynamic metabolic response. The fMRI analysis also has a finite dynamic response time, but it will be treated as incorporated in the MRK of the glial/hemodynamic response. The formal equations linking the processing stages in **Figure 1** are provided in **Table 1**.

DIRECT FORWARD MODELING APPROACHES

A few previous studies have made estimates of the effects of different neural models on the form of the BOLD response dynamics. For example, Mechelli et al. (2001) report a simulation study of the estimated regional cerebral blood flow (rCBF) and BOLD signals as a function of the duration, onset asynchrony, and relative amplitudes of two brief stimuli. They included a basic model of neuronal dynamics and varied one parameter of this model – the amplitude of a slow late transient – to show its effect on the simulated BOLD responses. This exercise constitutes an unvalidated forward model of the effect of only one parameter of a simulated neural response. However, what they offer as the analysis of the effects on the BOLD waveform is described (incomprehensibly) as “the BOLD parameter estimates,” which is actually a single-valued function with no specification of which parameter(s) is/are being estimated. Since no comparison is made with empirical data or their noise limitations, the fact that this study includes a proposed model for the neural dynamics does not qualify it as a validated procedure for estimating the neural population dynamics underlying the local BOLD signals (which is the goal of our study).

¹The term “kernel” is preferred because it is conceptualized as the theoretical response to an imaginary stimulus of infinitely short duration and unit area, termed a “Dirac pulse.”

Table 1 | Mathematical model of the linear and nonlinear operations involved in the generation of the BOLD signal from the input stimulus, $S(t)$.

Output	Generation logic	Features
Neural signal	$N_i(t) = \int [S(t) \otimes n_i(t)]$	Nonlinear transducer
Neural metabolic demand	$M_i(t) = \int [N_i(t) \otimes m(t)]$	Nonlinear transducer
Glial oxygen demand	$G(t) = \sum M_i(t) \otimes g(t)$	Multiple linear integrator
Hemodynamic response	$H(t) = G(t) \otimes h(t)$	Linear (slow)
Paramagnetic response	$Y(t) = H(t) \otimes p(t)$ $\approx \sum M_i(t) \otimes \text{MRK}(t)$	Linear (fast) Combines four linear stages

Buxton et al. (2004) extend their balloon model of the hemodynamic response leading to the BOLD signal by proposing a model of the neural response to account for the temporal non-linearity in BOLD responses as a function of duration. This neural response incorporates a slow subtractive inhibitory component to the net neural signal, which has the effect of producing a neural response consisting of an initial transient followed by a sustained plateau. Model responses for three kinds of stimuli – a single short pulse, two short pulses, and one long pulse, are offered as a demonstration of the properties of this model. As with Mechelli et al. (2001), no attempt is made to compare the model outputs with actual BOLD recordings, so the Buxton et al. (2004) study again does not qualify as a validated procedure for estimating the neural population dynamics underlying the local BOLD signals.

Both Mechelli et al. (2001) and Buxton et al. (2004) include parameters intended to account for the temporal non-linearity of short-duration responses (which do not fall linearly as response duration is reduced; Boynton et al., 1996; Birn et al., 2001). Both studies demonstrate the required lack of reduction in a single example of a short-duration response, but neither study provides a validation of either the waveform or the amplitude response function relative to empirical BOLD data. In principle, either model could provide a platform for such validation or the further estimation of the dynamics of the underlying neural population response, but they neither do so nor suggest procedures by which such estimation could be achieved.

STEPWISE INCREMENTAL ESTIMATION OF THE NEURAL DYNAMICS

Quantitative estimation of the temporal transients underlying BOLD responses can be achieved by the use of an appropriate set of temporal duration stimuli, such as the temporal summation study reported in Birn et al. (2001). Their stimuli consisted of counterphasing checkerboard patches with durations increasing in factors of 2 from 0.5–2 s (together with a 20-s duration stimulus). The key result was that the BOLD response amplitudes did *not* increase proportionately with stimulus duration. (A proportionate increase would be expected under a general linear model (GLM) of linear summation with a neural response of constant amplitude.) Instead, the amplitude showed little increase from the short durations, implying either a non-linear summation process or a disproportionate strength of neural signals for short-duration stimuli (**Figure 2**).

Bandettini and Ungerleider (2001) proposed a discrete approximation analysis for the derivation of the implied neural signal in human cortex under the assumption that the integration process underlying the generation of the BOLD response was indeed

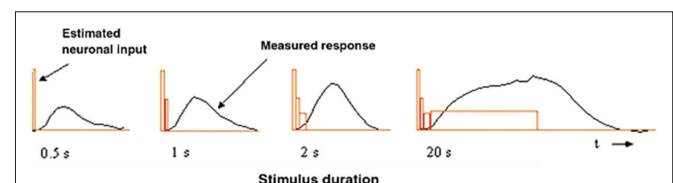


FIGURE 2 | Neuronal input amplitude estimation by a box-car approximation to the presumed neuronal signal (red rectangles) that accounts for the BOLD responses (black line) as a function of the duration of a counterphase checkerboard stimulus. Reproduced from Bandettini and Ungerleider (2001).

equivalent to a linear convolution operation. To derive a crude estimate of the neural signal, they took the difference between integrated BOLD response areas for each successive pair of stimulus durations as an estimate of the amplitude of the neural signal for the time interval defined by the difference in stimulus durations (Figure 2), based on the (unstated) assumption that the neural response is fast enough to follow the stimulus exactly. This discrete approximation approach implied that the neural signal underlying these BOLD responses has a pronounced early transient followed by a sustained phase of stable amplitude out to 20 s (red box functions, Figure 2). The details of this transient, however, remained unresolved.

NON-LINEAR DYNAMIC FORWARD OPTIMIZATION

In this section, we propose a more biophysically based development of the forward modeling approach that we term NDFO. Rather than simply characterizing the behavior of the BOLD waveform (Birn et al., 2001; Bandettini et al., 2002; d'Avossa et al., 2003; Fox et al., 2005; Grotz et al., 2009) or attempting to infer the potentially complex properties of the underlying neural mechanisms from the form of the BOLD response by deconvolution (Glover, 1999; Logothetis, 2002, 2003; Logothetis and Wandell, 2004), the concept of forward modeling is to start from the “cause” – the neural signal, instead from the “consequence” – the BOLD signal. Thus we incorporate into the model as much knowledge as possible about the likely neural substrate and then *optimize* the remaining details to best fit the BOLD waveform. This knowledge includes the known temporal properties of neural responses and non-linearities both of the population of neural responses to the stimulus and of the metabolic requirements of the neural processing that lead to the measurable BOLD response.

The dynamic forward modeling discussed so far is *linear* (see Boynton et al., 1996) in the sense that it assumes no non-linearity in the model linking the neural response to the BOLD response (or indeed in linking the original stimulus waveform to the BOLD response). It merely estimates the *gain* of the neural response for each stimulus duration, which can vary by a linear process. A more general form of forward modeling is to incorporate a variety of possible non-linearities into the structure of the model. The range of possible non-linearities at each stage of the model is so large that the approach is not feasible unless one incorporates Bayesian constraints in the modeling, based on the known biophysics of the neuronal response properties likely to underlie the BOLD activation (Logothetis and Wandell, 2004; see Table 1). The particular form of the model will depend on the specific knowledge of the population dynamics of the signal driving the hemodynamic response available from the literature at any given time.

The neural model that we investigate in the present version of the analysis is the sum of positive and a negative component based on delayed gamma functions convolved with the stimulus waveform, with the parametrization specified in Table 2. To illustrate the properties of the model, we analyze the effect of varying the inhibitory ratio implied by the negative component weight b , and the offset/onset gain ratio λ .

ANALYTIC FRAMEWORK FOR THE NEURAL TEMPORAL RESPONSE

The starting point for the NDFO modeling is a model of the neural signal, whose first effect in terms of the cascade of BOLD dynamics is to create a metabolic demand $G(t)$ in the neighboring glial

cells (see Figure 1). Gamma probability density functions have the analytic form $a \cdot t^{k-1} \cdot e^{-t/\tau} / \Gamma(k)$, where t is the time dimension, a is a scaling parameter, and k and τ are generic waveform parameters. (To equate the area to unity, the function is defined with $a = 1/\tau^k$, but for the present purposes a is a free optimization parameter.) They may be termed simply “gamma functions” to emphasize their analytic rather than statistical properties. In engineering the same function is known as the n -pole filter function (with $n = k$), and is used to describe the dynamics of a wide range of processes. For the present application, the temporal gamma function is assumed to have integer powers of k and corresponds to the solution of differential equations with real (non-imaginary) roots. The gamma function has the analytic advantage over many other functions, such as the Gaussian, that it is by definition causal because it has the value of 0 at $t = 0$, and is defined as 0 for $t < 0$ (i.e., the full specification is $y = k^{t-1} \cdot e^{-t/\tau}$ for $t < 0$; $y = 0$ for $t < 0$). Its shape progresses from highly asymmetric around the peak for small k to approximately Gaussian and symmetric for large k (see Figure 3).

A key feature of this formalism is that the peak latency is determined by the time constant, τ , and is proportional to the width (at half height) of the response peak, which can be estimated to good accuracy by the methods described in the next section.

NEURAL MODEL

A comprehensive model of the BOLD therefore requires an accurate model of the intracellular potential dynamics deriving from the sensory stimulation. Formally, we propose to use the full neural response model jointly specified in Eqs 1 and 2:

$$N_p(t) = \left[|N(t)|_+ + \lambda \cdot |N(t)|_- \right] e^{-\tau/\eta} + \varepsilon(t), \quad (1)$$

where ε is the net source of additive noise and the function is governed by the parameter set.

Table 2 | Non-linear forward model parameters.

a	Scaling constant (a fitting parameter but not a waveshape parameter)
k_n	Integer exponent governing the rising phase
τ_n	Time constant of falling phase in the neural response
Δt	Transmission delay before response onset
b	Sustained/transient ratio in the step response
η	Time constant of non-linear gain control in the neural response
λ	Ratio of offset to onset gain in the neural response

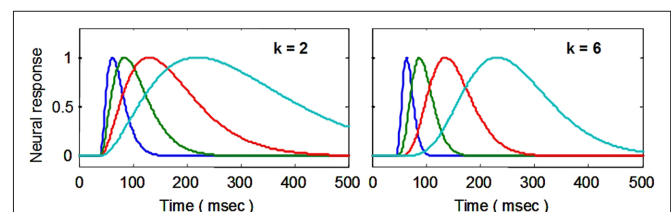


FIGURE 3 | Examples of delayed gamma function step responses with exponents of $k = 2$ (left) and 6 (right). Successive functions (colors) introduce a wide range of peak latencies for the neural signal estimates (with time constants τ increasing in factors of 2 and a fixed delay Δt of 40 ms).

The neural signal $N_p(t)$ in Eq. 1 is the sum of positive and a negative component based on delayed gamma functions convolved with the stimulus waveform:

$$N(t) = a \cdot S(t) \otimes n(t - \Delta t) \quad (2)$$

$$\text{where } N(t) = (1 - t) \cdot t^{k_n - 1} e^{-t/\tau_n} - \frac{b}{\tau_n} t^{k_n} e^{-t/\tau_n}$$

(Note the convention that time series functions are capitalized, impulse response kernels are lower case, and vectors are bold-face.) The neural impulse response expression $n(t)$ is set up so that its convolution with a step function is equivalent to the sum of a pure transient and a pure sustained component. In addition, the expression is specified with an additional transmission delay, Δt , that delays the response relative to the stimulus without affecting its waveshape. Thus, **Table 2** defines the parameters in vector $\mathbf{p}$ of this equation.

One issue that arises is how to measure the latency Δt of the delayed gamma functions of **Figure 3**. A simple derivation can show that the peak latency t_{peak} of these responses is specified by the expression $(k\tau_n + \Delta t)$. Thus if k and Δt , both of which are well-determined from neurophysiological studies in monkey cortex, are set to means of their Bayesian priors on this basis, the peak latency t_{peak} is determined from the value of τ , which can be accurately derived from the model optimization.

METABOLIC COUPLING

Since little is known about the glial dynamics of transmitter recovery, we will pursue two options as to their effects. One option is that the *metabolic demand* driving the BOLD response derives from the transmitter recovery cycle following the activation by an axonal spike. Since axonal spikes represent only the positive aspect of the intracellular voltage and since 90% of cortical synapses are excitatory (Shank and Aprison, 1979; Wang and Floor, 1994), the signal transmitted from one cortical stage to the next may be treated as a *half-wave rectified* version of the dynamic neural signal. This prediction is shown as the blue curves in **Figure 4Aa** (indexed in row/column notation), which is an overlay of the model estimates of the neural responses to stimulus pulses that double in duration from 8 ms to 16 s (eight doublings). For this example, the neural response has balanced excitation and inhibition, so even the prolonged pulses generate only an initial transient response, with the negative lobe at offset being thresholded out by the rectification. (Note that the local metabolic demand, $M_i(t)$, has the same time course in this model as the transmitter recovery from which it derives. The energetic processes required for the recovery to the initial state, however, form an oxygen-based chain of glial metabolic response, $G(t)$, that may have substantially slower time course at one or more stages.)

The other option is to consider the instantaneous metabolic demand of both excitatory and inhibitory cells, or both “on” and “off” cells, implying that the signal generating the metabolic demand is a *full-wave rectified* version of the intracellular voltage. This option is shown as the red curve in **Figure 4Aa**, whose spikes represent the instantaneous metabolic demand for the offset transient for each of the pulse duration doublings (which were thresholded out in the blue curve).

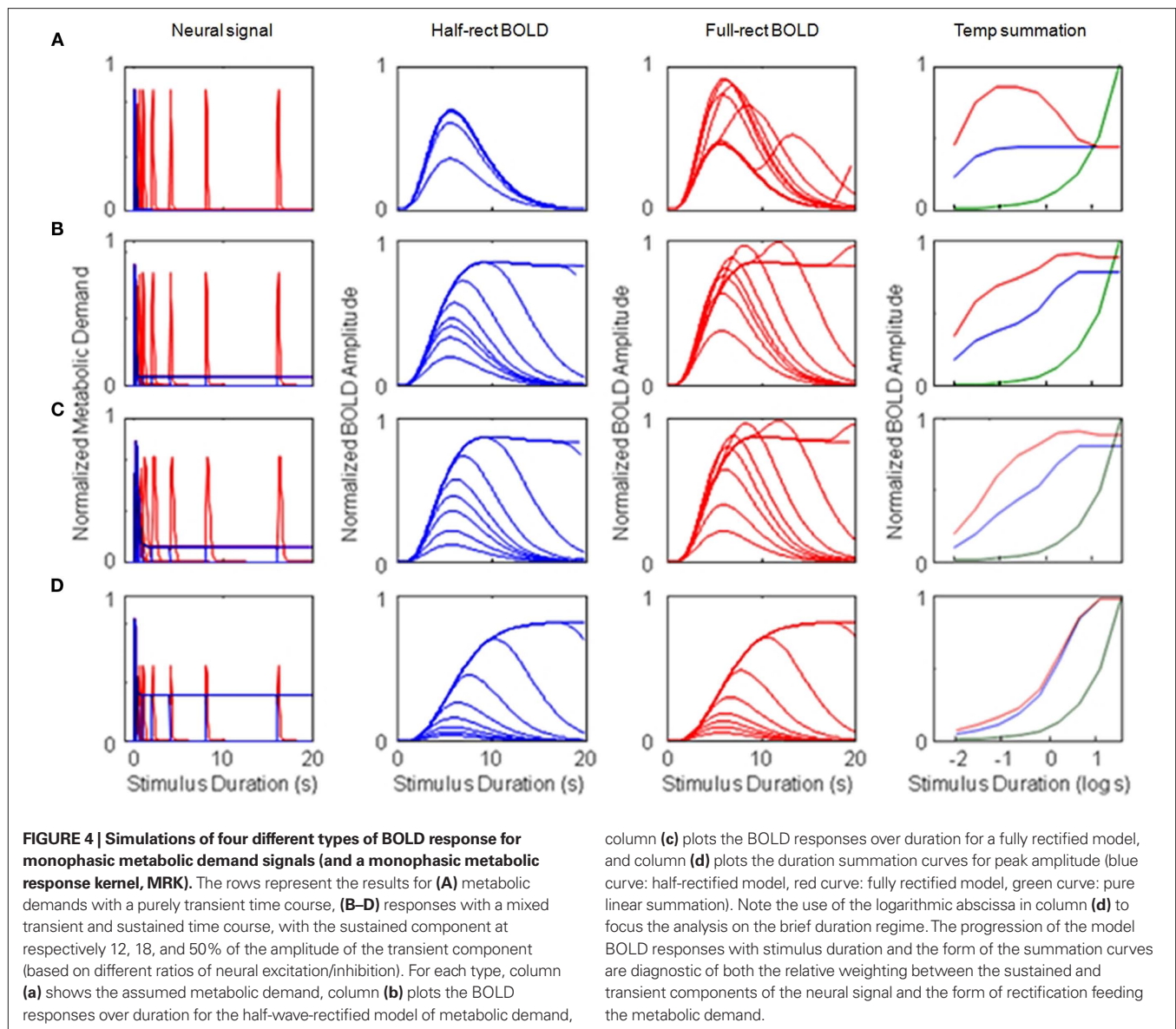
Having provided relevant variables for the linear and non-linear components of the neural response dynamics, we may now consider the issue of the metabolic coupling of the neural signal to generate the BOLD response. This coupling has been extensively modeled over the past two decades, with the best-known example being the Buxton–Friston balloon model and the most elaborated version being by Sotero and Trujillo-Barreto (2007, 2008). However, these models are not well-validated by empirical human studies (because they contain too many interdependent variables to allow the independent assessment of each separately).

CHARACTERISTICS OF THE MODEL

We may now evaluate the response to these two options for the non-linearity of the metabolic demand through the biophysical chain of the metabolic processes to the measured BOLD signal (**Table 1**). The first element in this chain is the astrocytes surrounding the neuron, which provide glucose to the neuron and replenish its supply by ATP metabolism fueled by oxygen from neighboring blood vessels. It may be emphasized that the astrocyte metabolic processes are as slow, relative to the intracellular signal dynamics, as are the processes of hemodynamic oxygen supply. The time constant of the astrocyte responses at the cell body is known to be of the order of several seconds (Kelly and van Essen, 1974; Filosa et al., 2004; Metea and Newman, 2006), and it is clear that these slow responses must mediate the hemodynamic component. However, at present too little is known of the dynamics of transmitter recovery and/or the non-linearities in the process to securely assign time constants to the astrocytic component relative to the hemodynamic component of the metabolic coupling. For the present example, we will therefore treat the entire chain from the metabolic demand to the magnetic resonance signal in the traditional fashion, as a unitary linear kernel. As stated above, this kernel is often termed the HRF, but in view of its likely substantial astrocyte contribution, we give it the more general term of the MRK. Our main goal is to estimate the properties of the neural signal processing, and it will be seen that there is sufficient information to provide a rich analysis of these properties, and to account for the empirical non-linearities of the BOLD signal, as long as the metabolic supply chain conforms to the linearity assumption. (As more information becomes available, non-linear aspects of the metabolic coupling may readily be incorporated into the analysis we propose.)

The development of the peak amplitude in the BOLD temporal summation series is shown as the blue summation curve in **Figure 4Ad**. The critical point of this plot is that the asymptotic corner of this summation curve occurs at 40 ms, which is the value of the time constant assumed for the neural signal in this example. Thus, the form of the BOLD amplitude summation series (**Figure 4**, column d) provides a direct empirical estimate of the time constant of neural integration down to the millisecond range. There is no limit in principle to the temporal resolution that can be achieved by this methodology since it is estimated from the *amplitude* variation of the BOLD signal as a function of stimulus duration, not from its temporal aspects.

The reduction in peak amplitude is captured in the red temporal summation curve of **Figure 4Ad**, showing a reduction by a factor of two for long durations. (The green curve in **Figure 4Ad** represents



the values expected for fully proportional linear summation of the energy in the stimulus pulse; it is an accelerating curve due to the logarithmic abscissa.) The case of the full-rectification model of the metabolic demand is shown in **Figure 4Ac**. Here the second neural response peak (i.e., that from the stimulus offset) plays a key role in varying the BOLD waveform, which first extends in time and then shows a two-peaked structure with reduced amplitude for long-duration stimuli.

Figure 4Ba shows the *half-* and *fully rectified* version of the metabolic demand to the same pulse duration series, where the neural inhibition is now assumed to be reduced in energy by 1.5% relative to the excitation. This small imbalance is magnified by the convolution with the sustained stimulus, and thus it results in a sustained component that is 12% of the amplitude of the initial transient (blue curve in **Figure 4Ba**) and then into an almost fully sustained set of BOLD response functions (**Figure 4Bb**, compare to **Figure 4Ab**). Thus, the *form* of the BOLD response functions

can be strongly diagnostic of even slight variations in the properties of brief *neural* signals. Moreover, the nature of the metabolic demand function (half- or fully rectified) has a big impact on the form of the BOLD response, determining whether or not an offset peak occurs at the tail of the responses even when they are sustained (**Figure 4Bc**). Such a peak has been reported in some studies but is not always evident. Thus it remains an empirical question to what extent rectification is representative of BOLD waveforms; intermediate forms of the rectification model are required to capture the empirical properties in detail.

Note that the amplitude series in **Figures 4Bb,c** show bands of denser packing of the functions, where the amplitude changes were not spaced in proportion to the doublings of stimulus duration. Viewed in terms of the sequence of BOLD waveforms in **Figures 4Bb,c**, the regions of dense packing form an intermediate “shelf” or partial asymptote in peak amplitude summation plots of **Figure 4Bd**. It is again evident that the onset of this intermediate

shelf in the summation curve corresponds to the integration time of the underlying neural signal, while the second asymptote at higher amplitude corresponds to the ~ 5 s integration time of the MRK (HRF). Accurate measurement of such summation functions can therefore provide *discriminative* characteristics that, when interpreted through the non-linear model structure, can provide estimates of *both* the neural and the metabolic time constants in the neural-to-BOLD signal chain.

This point is emphasized by the response set in row c of **Figure 4**, which probes the effect of varying the *time constant* of the neural transient. The key difference from the parameters used in row b of **Figure 4**, is that the neural time constant was doubled from 50 to 100 ms (and the excitation/inhibition imbalance was also increased to 7% to maintain the same form of offset peak). It is evident that (i) the summation curve (**Figure 4Cd**) takes a measurably different form, and that (ii) the accuracy of estimation of the neural time constant is limited not by the BOLD time constant but only by the variability of the BOLD amplitude measures. For example, this analysis shows that the neural time constant is estimable to within about 0.1 log units if the BOLD response functions can be measured to an achievable accuracy of about 10%.

The final case of duration summation analyses (**Figure 4Dd**) shows the NDFO predictions of increasing the excitation/inhibition imbalance of the neural response to 50%, illustrative of a system that is predominantly sustained in nature. Under these conditions, the impact of the initial transient becomes essentially negligible, and the summation curves (**Figure 4Dd**) become indistinguishable from proportional summation (i.e., they run parallel to the green curve). This manipulation illustrates that the power of the NDFO analysis depends on the neural processing being predominantly transient, and that the properties of the underlying neural mechanisms would not be accessible to this form of analysis in predominantly sustained systems. Luckily, however, the well-established deviation from proportionality for short-duration stimuli implies that the neural system is, in practice, predominantly transient and is therefore amenable to this form of NDFO analysis.

VALIDATION OF THE FORWARD MODEL ANALYSIS

To validate the model framework described in the previous sections in a specific implementation, and to determine the sensitivity of the model parameters to experimental noise sources, we conducted a specific BOLD fMRI experiment.

METHODS

Ethics statement

This research was approved by the institutional review board of the University of California, San Francisco, with informed consent obtained for the fMRI study.

Stimulus presentation

The stimuli were spatial arrays of Gabor patches scaled to maximally stimulate retinotopic cortices at all eccentricities. They were presented in durations from a single frame of 16.67 ms duration in factors of 2 to 1.067 s (**Figure 5A**) in a fully balanced m-sequence design. The durations were presented with full counterbalancing by

the m-sequence design. M-sequences are pseudorandom sequences that fully counterbalance the order of stimulus presentation up to a memory length of m-sequential items (Golomb, 1967; Sutter, 1987, 1991; Buracas and Boynton, 2002). They therefore specify the ideal order of presentation of stimuli given that the memory length of the system is known. The m-sequence design is constrained by the fMRI modality, since its analysis is inherently slower than for the ERP. The interstimulus interval was set at 4 s, as we have found that this is an optimal rate for fMRI studies. The typical BOLD response lasts for about 16 s, so this is the length of time over which the response sequence needs to be balanced. Quantitatively, with a sampling time of $TR = 2$ s and an interstimulus interval of 4 s, we used an m-sequence that balanced all combinations of eight stimuli over a memory duration of 16 s ($= 4$ samples).

fMRI data collection

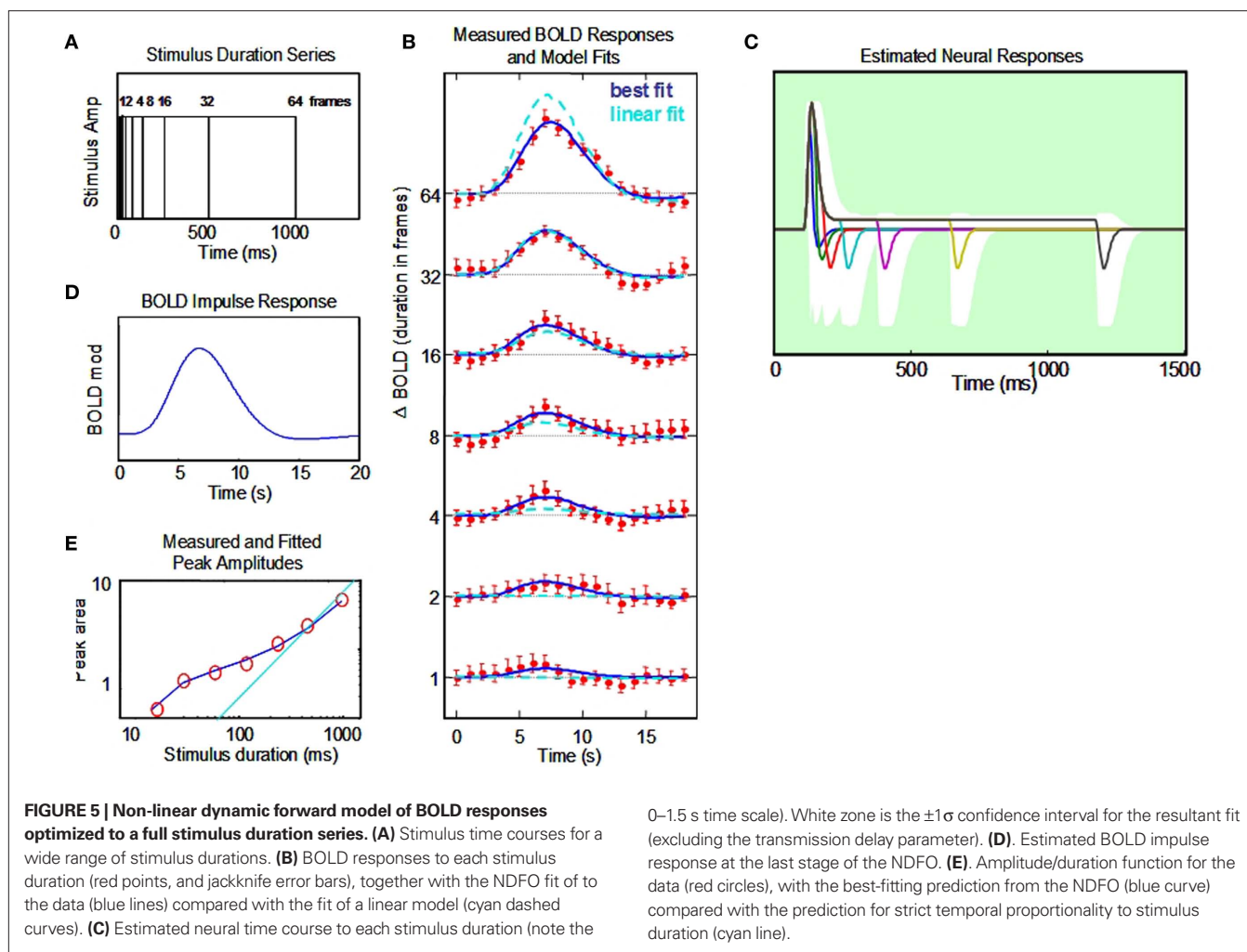
Functional MRI scanning was conducted on a Siemens Trio 3 T scanner equipped with eight-channel SENSE parallel imaging capability. In targeting the specific activation sites, it is important to avoid artifactual spread of the activation by partial-voluming across cortex and between blood vessels and cortex on the two sides of a sulcus. Such artifactual spread may account for much of the apparently distributed activation in previous studies with large voxels, and is almost never discussed in relation to studies of the activation patterns for visual processing tasks.

To avoid partial-voluming with the blood vessel overlying the cortex, we make the voxels small enough to lie strictly within the cortical gray matter. For a cortical thickness of 2.5 mm, a sampling analysis shows that the $\sim 3^3$ mm voxels in a typical retinotopy scan have an average of more than half (60%) of their volume *outside* the cortex, and even across to the other side of a sulcus. (The situation is obviously far worse if the data are averaged to 4^3 mm effective voxel size. If, however, the voxels are reduced to 2^3 mm, a continuous sheet of voxels could be found to lie almost entirely *within* the cortex (an average of 80% isolation of each sample volume).

Our current recording parameters achieve such resolution with an in-plane matrix size of 96 at 2^3 mm isotropic resolution by using $TR = 2$ s (with $TE = 27$ ms, flip angle $= 90^\circ$), interleaved by staggering the event timings to give an effective temporal resolution of 1 s for the BOLD temporal waveform (see Appendix for details). The 4 cm depth provided sufficient coverage for the posterior occipital lobe in each hemisphere. Activations were analyzed only for voxels lying within the cortical gray matter, as defined by an anatomical segmentation algorithm). The voxels were selected as those lying within the calcarine sulcus, and were therefore restricted to primary visual cortex (V1).

Error bars

To obtain both the individual responses and the standard errors required for the maximum entropy estimation procedure, we employ a novel use of the jackknife procedure specified by Efron and Tibshirani (1993). In brief, to determine the error variance for each repeat in an experiment with $i = 1, 2, \dots, n$ repeats of a given stimulus condition, $n - 1$ jackknife estimates for each of the event types in the design matrix are derived by re-running the analysis with each repeat of the chosen event removed in succession from the design matrix.



Under the jackknife procedure, the standard error of the mean (SEM) for each datum point of the response matrix $X(k, t)$ for each event type k at each time point t is calculated from the i th-deleted pseudovalues according to

$$\text{SEM} = \sqrt{\frac{\sum_{i=1}^n \frac{\tilde{X}_i - \Sigma \tilde{X}_i / n}{n(n-1)}}{n(n-1)}} \quad (3)$$

where $\tilde{X}_i = n\tilde{A} - (n-1)\tilde{A}_i$, with $\tilde{A}$ and $\tilde{A}_i$ being the average and the i th-deleted average vectors, respectively.

RESULTS

BOLD RESPONSES

To validate our conceptual breakthrough for the estimation of neural response dynamics from BOLD response measurements we performed a computational analysis of the implications of the BOLD fMRI data collected on a typical human subject with corrected to normal vision, a temporal modulation sensitivity function within normal limits, typical event-related potentials to contrast reversal (onset latency of 52 ms and N1 latency of 115 ms) and a typical BOLD response time course as evident from Figure 5 and a number of previous retinotopic and localizer scans. The analy-

sis was conducted in a full-range temporal summation paradigm (Figure 5A) and provides not only the estimated neural response waveform but confidence limits on the waveform parameters that are sufficiently narrow to indicate that the results are within the expected range for early cortical response dynamics.

The responses from primary visual cortex (~250 voxels) for a 1-h session from a typical subject are shown in Figure 5B. They show a progressive growth in amplitude with stimulus duration from the shortest to the longest, but note that the amplitudes do not increase by a factor of two (the linear GLM prediction; cyan dashed curves in Figure 5B) as duration is increased. We therefore need a non-linear model to understand the processing mechanism for these effects.

The data have thus been fit through our NDFO procedure of a linear convolution of the stimuli from durations from 16.67 ms to 1.067 s in factors of 2 (Figure 5A) with each neural component kernel to generate the neural signals at each duration (Figure 5C), a non-linear coupling to generate the component metabolic demands, a linear summation over the component demands to generate the net metabolic demand, and a linear convolution with a MRK (Figure 5D) to generate the ultimate BOLD signal prediction (Figure 5B). Figure 5E is a comparison of the amplitudes of

the fitted model waveforms with those of the data, as analyzed in Section “Discussion.” Optimization of the BOLD prediction (blue curves in **Figure 5B**) to the data determines the temporal features of the requisite neural signals making up each waveform, allowing us to specify the predominant time constants for the underlying neural signals. These features are defined by the following model population parameters for the net neural signal throughout each studied cortical region: transmission delay, gamma exponent, recovery time constant, sustained/transient ratio, sustained decay, and off/on ratio (six waveform parameters). The amplitude, transient integration time, and sustained/transient ratio are specified to worthwhile precision (see Appendix). The off/on ratio parameter controlling the relative amplitude of the off-responses is not well-constrained for the duration-range used in this data set (**Figure 5**), but we note that this parameter could also be brought under tighter control by adding stimuli of longer duration (e.g., 8 s). This approach should readily provide tight constraints on the off/on ratio because the BOLD activity corresponding to the off-responses is pushed to a time period that is essentially independent of that for the on response. This kind of approach to the off transient had been taken successfully by Fox et al. (2005) and Lingnau et al. (2009), for example.

DISCUSSION

Our novel analysis of the neural signal underlying the BOLD response waveforms to a full temporal stimulus series constitutes a form of “temporal microscope” for the neural signals in the cortex generating the recorded BOLD waveforms. Thus, this study illustrates how the neural waveform parameters can be estimated from the temporal summation series to their native temporal resolution. The estimated neural waveform for the present dataset in **Figure 5C** has a strong brief transient with a width at half height of 55 ms, followed by a sustained response of about 1/10th of the peak transient response and a reduced, but rectified, offset response at the end of each stimulus. In each respect, these response properties match the corresponding neural response properties recorded in the cortex of other species. A useful survey of the dynamics of the unitary intracellular responses to single synaptic action potentials is provided by Thomson and Lamy (2007). They tabulate data from a large number of studies of the excitatory post-synaptic potential (EPSP) time course, 54 of which have usable data on the exponential time constant of decay. These are analyzed in Section “Appendix” to show that the typical EPSP time constant has a standard deviation of ± 20 ms. This kind of analysis can provide the Bayesian constraints on the modeling, although we note that the typical afferent volley has a temporal spread of about ± 20 ms that must be convolved with the EPSP time constant to obtain an estimate of the metabolic demand generated by the typical synaptic activation. Thus, the net time course of the neural transients is expected to have a standard deviation of the order of 28 ms, or a full width at half height of ~ 50 ms. We are not aware of comparable meta-analysis for monkey cortical responses, but this value is clearly in line with the published examples of V1 transients from this species (such as those of Hegdé and van Essen, 2004).

It must be emphasized that the transient time-constant analysis relies on the deviation from the linear model of proportional responses (indicated by the cyan line in amplitude plot of

Figure 5E). Such deviations can be accurately determined because they are based on the integrated response amplitude at short durations. The accuracy of the model fit does not depend on this linearity assumption, but could be fit equally well to a non-linear temporal integration behavior if that behavior were specifiable for typical neural populations. To our knowledge, however, neural signals exhibit linear temporal integration at this time scale (e.g., Koch, 2005), and the primary non-linearity is the inability of the spike rate to fall below 0, coupled with the presence of both ON and OFF type neurons, which result in a fully rectified spike-rate response for the typical population of a cortical voxel. Just how the spike rate is coupled to the metabolic demand, in terms of its rectification behavior, has not yet been determined, which is why we provide an example analysis in **Figure 4** of how the determination of one form of non-linearity can be made in the framework of the present analysis.

Similarly, as knowledge evolves, more complex models of the neural/metabolic coupling and the hemodynamic response could be easily incorporated in our model. These all represent Bayesian information that, if well-established, can be used to refine the model structure and enhance the fitting process when available. However, our understanding of the literature is that (a) the first-order specification of the neural/BOLD coupling is well approximated by a linear response kernel, and (b) that estimates of the second-order effects are contaminated by the assumption that they are purely hemodynamic, and have not taken into account potential and actual non-linearities in the neural signals at the time scale of the BOLD signal. Thus, despite the best efforts of the proponents of elaborated hemodynamic modeling, there is no secure information about the non-linearities of this process for the human brain *in vivo*. Consequently, we have tried to redress the balance by considering the effects plausible non-linearities in the neural population responses to standard types of stimulus presentation. For clarity in this enterprise, we have assumed that the metabolic response is both linear and monophasic, illustrating a variety of BOLD response properties that could arise from such neural non-linearities. We are not claiming to have proven that these BOLD properties are entirely determined at the neural level (which would be a vast enterprise, far beyond the scope of one paper), but that claims that they are purely properties of the vascular hemodynamics *per se* are suspect until they are replicated in paradigms that remove the neural component of the system [such as stimulation of the hemodynamic response by direct infusion of nitric oxide (NO) in the vicinity of the blood vessels. This experiment has apparently not been attempted, although Burke and Bührle (2006) showed that suppression of the NO with application of the NO synthase inhibitor 7-nitroindazole completely abolished the BOLD response while only marginally affecting the local field potentials, establishing the role of NO in neurovascular coupling.]

In particular, the non-linearity of the transient responses at the offset of the neural response could be positive (rectifying) rather than negative (linear), and could in either case show reduced amplitude relative to the onset response (adaptive gain control). In this example of the NDFO analysis, the offset responses are much smaller than the positive responses at stimulus onset. This analysis demonstrates that estimation of the neural response dynamics for each stimulus type is therefore not a Herculean task

requiring extreme levels of signal/noise ratio, but is well within the capability of one session of normal fMRI methodology using, however, the appropriate combination of experimental design and theoretical analysis.

We note that a technique with a philosophy similar to the present approach has been successfully applied to the estimation of net spatial receptive field structure of small cortical regions by Dumoulin and Wandell (2008), although they used a linear rather than the full non-linear model of the sequence of processes. Their spatial estimates were based on a model of the temporal signal to be expected as a stimulus swept across each defined point on the retina. Like us, they take the temporal stimulus waveform, convolve it with a spatiotemporal model of the response of the underlying neural population, and then with a model of the MRK (HRF) to provide a forward model of the temporal BOLD response that is optimized to the measured BOLD response at each cortical location. Our approach takes the temporal analysis several steps further toward biological plausibility, and focuses on the temporal rather than spatial aspect of the neural population response, but the general philosophy of analyzing interesting parameters of the neural response through a forward model of the BOLD response is a similar one.

REFERENCES

- Aguirre, G. K., Zarahn, E., and D'Esposito, M. (1998). The variability of human BOLD hemodynamic responses. *Neuroimage* 8, 360–369.
- Azouz, R., and Gray, C. M. (1999). Cellular mechanisms contributing to response variability of cortical neurons in vivo. *J. Neurosci.* 19, 2209–2223.
- Bandettini, P. A., and Ungerleider, L. G. (2001). From neuron to BOLD: new connections. *Nat. Neurosci.* 4, 864–866.
- Bandettini, P. A., Birn, R. M., Kelley, D., and Saad, Z. S. (2002). Dynamic nonlinearities in BOLD contrast: neuronal or hemodynamic? *Elsevier Excerpta Medica Int. Congr. Ser.* 1235, 73–85.
- Barth, M., and Norris, D. G. (2007). Very high-resolution three-dimensional functional MRI of the human visual cortex with elimination of large venous vessels. *NMR Biomed.* 20, 477–484. doi: 10.1002/nbm.1158
- Bellgowan, P. S., Saad, Z. S., and Bandettini, P. A. (2003). Understanding neural system dynamics through task modulation and measurement of functional MRI amplitude, latency, and width. *Proc. Natl. Acad. Sci. U.S.A.* 10, 1415–1419.
- Bevington, P. R., and Robinson, D. K. (2003). *Data Reduction and Error Analysis for the Physical Sciences*. New York, NY: McGraw-Hill.
- Birn, R. M., Saad, Z. S., and Bandettini, P. A. (2001). Spatial heterogeneity of the nonlinear dynamics in the fMRI BOLD response. *Neuroimage* 14, 817–826.
- Boynton, G. M., Engel, S. A., Glover, G. H., and Heeger, D. J. (1996). Linear systems analysis of functional magnetic resonance imaging in human V1. *J. Neurosci.* 16, 4207–4221.
- Buracas, G. T., and Boynton, G. M. (2002). Efficient design of event-related fMRI experiments using m-sequences. *Neuroimage* 16, 801–813.
- Burke, M., and Bühlre, C. (2006). BOLD response during uncoupling of neuronal activity and CBF. *Neuroimage* 32, 1–8.
- Buxton, R. B., Uludağ, K., Dubowitz, D. J., and Liu, T. T. (2004). Modeling the hemodynamic response to brain activation. *Neuroimage* 23 (Suppl 1), S220–S233.
- Carandini, M., and Ferster, D. (1997). A tonic hyperpolarization underlying contrast adaptation in cat visual cortex. *Science* 276, 949–952.
- Carandini, M., and Ferster, D. (2000). Membrane potential and firing rate in cat primary visual cortex. *J. Neurosci.* 20, 470–484.
- Celebi, S., and Principe, J. C. (1995). Parametric least squares approximation using gamma bases. *IEEE Trans. Signal Process.* 43, 781–784.
- Chatton, J. Y., Pellerin, L., and Magistretti, P. J. (2003). GABA uptake into astrocytes is not associated with significant metabolic cost: implications for brain imaging of inhibitory transmission. *Proc. Natl. Acad. Sci. U.S.A.* 100, 12456–12461.
- Chen, S. (2006). Signal processing. *Eur. Trans. Telecommun.* 17, 99–110.
- Cohen, J. (1992). A power primer. *Psychological Bulletin* 112, 155–159.
- d'Avossa, G., Shulman, G. L., and Corbetta, M. (2003). Identification of cerebral networks by classification of the shape of BOLD responses. *J. Neurophysiol.* 90, 360–371.
- De Vries, B., and Principe, J. C. (1992). The gamma model: a new neural model for temporal processing. *Neural Netw.* 5, 565–576.
- de Zwart, J. A., Silva, A. C., van Gelderen, P., Kellman, P., Fukunaga, M., Chu, R., Koretsky, A. P., Frank, J. A., and Duyn, J. H. (2005). Temporal dynamics of the BOLD fMRI impulse response. *Neuroimage* 24, 667–677.
- Dumoulin, S. O., and Wandell, B. A. (2008). Population receptive field estimates in human visual cortex. *Neuroimage* 39, 647–660.
- Efron, B., and Tibshirani, R. J. (1993). *An Introduction to the Bootstrap*. New York: Chapman & Hall.
- Farwell, V. T., and Prentice, R. L. (1977). A study of distributional shape in life testing. *Technometrics* 19, 69–75.
- Filosa, J. A., Bonev, A. D., and Nelson, M. T. (2004). Calcium dynamics in cortical astrocytes and arterioles during neurovascular coupling. *Circ. Res.* 95, 73–81.
- Fox, M. D., Snyder, A. Z., Barch, D. M., Gusnard, D. A., and Raichle, M. E. (2005). Transient BOLD responses at block transitions. *Neuroimage* 28, 956–966.
- Friston, K. J. (1997). Transients, metastability, and neuronal dynamics. *Neuroimage* 5, 164–171.
- Friston, K. J., Fletcher, P., Josephs, O., Holmes, A., Rugg, M. D., and Turner, R. (1998). Event-related fMRI: characterizing differential responses. *Neuroimage* 7, 30–40.
- Friston, K. J., Mechelli, A., Turner, R., and Price, C. J. (2000). Nonlinear responses in fMRI: the Balloon model, Volterra kernels, and other hemodynamics. *Neuroimage* 12, 466–477.
- Glover, G. (1999). Deconvolution of impulse response in event-related BOLD fMRI. *Neuroimage* 9, 416–429.
- Golomb, S. W. (1967). *Shift Register Sequences*. San Francisco: Holden-Day.
- Grotz, T., Zahneisen, B., Ella, A., Zaitsev, M., and Hennig, J. (2009). Fast functional brain imaging using constrained reconstruction based on regularization using arbitrary projections. *Magn. Reson. Med.* 62, 394–405.
- Handwerker, D. A., Ollinger, J. M., and D'Esposito, M. (2004). Variation of BOLD hemodynamic responses across subjects and brain regions and their effects on statistical analyses. *Neuroimage* 21, 1639–1651.
- Harel, N., Lin, J., Moeller, S., Ugurbil, K., and Yacoub, E. (2006). Combined imaging-histological study of cortical laminar specificity of fMRI signals. *Neuroimage* 29, 879–887.
- Hegd , J., and van Essen, D. C. (2004). Temporal dynamics of shape analysis in macaque visual area V2. *J. Neurophysiol.* 92, 3030–3042.
- Hegd , J., and Van Essen, D. C. (2006). Temporal dynamics of 2D and 3D shape representation in macaque visual area V4. *Vis. Neurosci.* 23, 749–763.

CONCLUSION

The conceptualizations and techniques introduced in this paper provide an analytic capability to resolving the timing and neural signal estimation underlying the BOLD waveforms recorded throughout the cortex. Any such attempt must be based on a model of the known neural dynamics of the neural populations underlying the BOLD metabolic signal generation, which may be progressively refined as more information becomes available about both neural response characteristics and the metabolic cascade. Given adequate signal/noise ratio, the present analysis shows that it is possible to develop approaches that overcome the temporal limitations of BOLD signal and are able to reveal the relevant properties of the underlying neural signals down to their native temporal resolution. In combination, these approaches represent a notable advance in the capabilities of the fMRI technology, providing a direct linkage between the live assessment of the functioning brain and the direct neurophysiological recordings in other species.

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- Heinzer, S., Krucker, T., Stampanoni, M., Abela, R., Meyer, E. P., Schuler, A., Schneider, P., and Müller, R. (2006). Hierarchical microimaging for multiscale analysis of large vascular networks. *Neuroimage* 32, 626–636.
- Henson, R. N., Price, C. J., Rugg, M. D., Turner, R., and Friston, K. J. (2002). Detecting latency differences in event-related BOLD responses: application to words versus nonwords and initial versus repeated face presentations. *Neuroimage* 15, 83–97.
- Hoge, R. D., Franceschini, M. A., Covolan, R. J., Huppert, T., Mandeville, J. B., and Boas, D. A. (2005). Simultaneous recording of task-induced changes in blood oxygenation, volume, and flow using diffuse optical imaging and arterial spin-labeling MRI. *Neuroimage* 25, 701–707.
- Kelly, J. P., and van Essen, D. C. (1974). Cell structure and function in the visual cortex of the cat. *J. Physiol.* 238, 515–547.
- Koch, C. (2005). *Biophysics of Computation: Information Processing in Single Neurons*. New York: Oxford University Press.
- Likova, L. T., and Tyler, C. W. (2007). Instantaneous stimulus paradigm: cortical network and dynamics of figure-ground organization. *Proc. SPIE* 6492, 64921E.
- Lin, F. H., Witzel, T., Mandeville, J. B., Polimeni, J. R., Zeffiro, T. A., Greve, D. N., Wiggins, G., Wald, L. L., and Belliveau, J. W. (2008). Event-related single-shot volumetric functional magnetic resonance inverse imaging of visual processing. *Neuroimage* 42, 230–247.
- Lingnau, A., Ashida, H., Wall, M. B., and Smith, A. T. (2009). Speed encoding in human visual cortex revealed by fMRI adaptation. *J. Vision* 9, 3.1–14.
- Logothetis, N. K. (2002). The neural basis of the BOLD fMRI signal. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* 357, 1003–1037.
- Logothetis, N. K. (2003). The underpinnings of the BOLD functional magnetic resonance imaging signal. *J. Neurosci.* 23, 3963–3971.
- Logothetis, N. K., and Wandell, B. A. (2004). Interpreting the BOLD signal. *Annu. Rev. Physiol.* 66, 735–769.
- Magistretti, P. J. (2009). Role of glutamate in neuron-glia metabolic coupling. *Am. J. Clin. Nutr.* 90, 875S–880S.
- Magistretti, P. J., and Pellerin, L. (1999). Cellular mechanisms of brain energy metabolism and their relevance to functional brain imaging. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* 354, 1155–1163.
- Martindale, J., Mayhew, J., Berwick, J., Jones, M., Martin, C., Johnston, D., Redgrave, P., and Zheng, Y. (2003). The hemodynamic impulse response to a single neural event. *J. Cereb. Blood Flow Metab.* 23, 546–555.
- Mechelli, A., Price, C. J., and Friston, K. J. (2001). Nonlinear coupling between evoked rCBF and BOLD signals: a simulation study of hemodynamic responses. *Neuroimage* 14, 862–872.
- Menon, R. S., Luknowsky, D. C., and Gati, J. S. (1998). Mental chronometry using latency-resolved functional MRI. *Proc. Natl. Acad. Sci. U.S.A.* 95, 10902–10907.
- Metea, M. R., and Newman, E. A. (2006). Glial cells dilate and constrict blood vessels: a mechanism of neurovascular coupling. *J. Neurosci.* 26, 2862–2870.
- Meyer, E. P., Ulmann-Schuler, A., Staufenbiel, M., and Krucker, T. (2008). Altered morphology and 3D architecture of brain vasculature in a mouse model for Alzheimer's disease. *Proc. Natl. Acad. Sci. U.S.A.* 105, 3587–3592.
- Nowak, L. G., and Bullier, J. (1997). "The timing of information transfer in the visual system," in *Cerebral Cortex and Extrastriate Cortex in Primates*, Vol. 12, eds K. Rockland, J. Kaas, and A. Peters (New York, NY: Plenum Press), 205–241.
- Shank, R., and Aprison, M. (1979). "Biochemical aspects of the neurotransmitter function of glutamate," in *Glutamic Acid: Advances in Biochemistry and Physiology*, eds L. J. Filer, Jr. S. Garattini, M. R. Kare, W. A. Reynolds, R. J. Wurtman (New York, NY: Raven), 139–150.
- Shao, X. M. (1997). Parametric survival analysis for gating kinetics of single potassium channels. *Brain Res.* 770, 96–104.
- Shmuel, A., Augath, M., Oeltermann, A., and Logothetis, N. K. (2006). Negative functional MRI response correlates with decreases in neuronal activity in monkey visual area V1. *Nat. Neurosci.* 9, 569–577.
- Sotero, R. C., and Trujillo-Barreto, N. J. (2007). Modelling the role of excitatory and inhibitory neuronal activity in the generation of the BOLD signal. *Neuroimage* 35, 149–165.
- Sotero, R. C., and Trujillo-Barreto, N. J. (2008). Biophysical model for integrating neuronal activity, EEG, fMRI and metabolism. *Neuroimage* 39, 290–309.
- Stacy, E. W. (1962). A generalization of the gamma distribution. *Ann. Math Stat.* 33, 1187–1192.
- Sun, F. T., Miller, L. M., and D'Esposito, M. (2005). Measuring temporal dynamics of functional networks using phase spectrum of fMRI data. *Neuroimage* 28, 227–237.
- Sutter, E. E. (1987). "A practical non-stochastic approach to nonlinear time-domain analysis," in *Advanced Methods in Physiological System Modelling*, Vol. 1, ed. V. Z. Marmarelis (Los Angeles: University of Southern California Press), 303–315.
- Sutter, E. E. (1991). The fast m-transform: a fast computation of cross-correlation with binary m-sequences. *SIAM J. Comput.* 20, 686–694.
- Thomson, A. M., and Lamy, C. (2007). Functional maps of neocortical local circuitry. *Front. Neurosci.* 1:1, 19–42.
- Thompson, J. K., Peterson, M. R., and Freeman, R. D. (2003). Single-neuron activity and tissue oxygenation in the cerebral cortex. *Science* 299, 1070–1072.
- Thompson, J. K., Peterson, M. R., and Freeman, R. D. (2004). High-resolution neurometabolic coupling revealed by focal activation of visual neurons. *Nat. Neurosci.* 7, 919–920.
- Thompson, J. K., Peterson, M. R., and Freeman, R. D. (2005). Separate spatial scales determine neural activity-dependent changes in tissue oxygen within central visual pathways. *J. Neurosci.* 25, 9046–9058.
- Wang, Y., and Floor, E. (1994). Dynamic storage of glutamate in rat brain synaptic vesicles. *Neurosci. Lett.* 180, 175–178.

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APPENDIX

THE CHAIN OF PROCESSING

The primary source of the BOLD signal may be estimated from studies of direct oxygen-level sensing in cortex (Thompson et al., 2003, 2004, 2005). These studies show that there are two identifiable components to the oxygen response to local luminance stimuli, a reduction in local oxygen level or negative response to the X and an increase in oxygen level or positive response to the Y. The spatial ranges of these responses were very different, with the negative response having a range of 50–100 μm from the sensor tip while the positive response had a range of 250–1000 μm . Since the distance from any neuron to its supporting arteriole is no greater than about 100 μm (Heinzer et al., 2006; Meyer et al., 2008), it seems clear that the range of the larger component, at least, is mediated largely by the spread of the neural processes. In fact, their model is based on the concept that “changes in tissue oxygen are composed of two response components (one positive and one negative) with magnitudes determined by neural activity on separate spatial scales” (Thompson et al., 2005). Since the typical spread of the more extensive component is about 10 times that of the more restricted component, the implication is that the volume of neural tissue expressing the former is about 100 times greater than of the latter. Any imaging technique, such as fMRI, with a spatial resolution of 1 mm or more is thus likely to be heavily dominated by the positive component.

Logothetis and colleagues (Logothetis, 2002, 2003; Logothetis and Wandell, 2004; Shmuel et al., 2006) have made the case that the most likely basis of the metabolic demand driving the BOLD signal is the energetic load deriving from the total conductance changes in the post-synaptic membrane generated by a range of processes consequent on transmitter release at the synaptic inputs to each neuron. The summed metabolic demand in the nexus of active cortical neurons adjacent to a capillary forms the drive for the metabolic response in that region of cortex. These authors argue that the transmitter release is tightly coupled to the activation of the post-synaptic receptors on the recipient cell membrane and consequently to the energetic demands of the membrane receptor activation and to a lesser extent to the subsequent recycling of the transmitter molecules. The majority of these energetic demands are met by glycolysis of glutamate to glutamine in the neighboring astrocytes (Shank and Aprison, 1979; Wang and Floor, 1994). The glutamine is then taken up by the neurons for reconversion to glutamate for use as a transmitter, releasing energy within the neuron in the process.

Since cells are predominantly linear summators of the excitatory and inhibitory transmitter release across their synaptic population, it seems to follow that the energetic demand driving the BOLD signal is most closely coupled to the net transmitter signal impinging on the cells, and hence to intracellular potential in the cells.

We treat the *neural responses* within each voxel as generated by sets of homogeneous populations with similar signal waveforms $N_i(t)$ within each population (Figure 1 in main text). Each neural response then generates a *local metabolic demand* $M_i(t)$ that may have a non-linear relationship to the neural signal waveform. The *integrated metabolic demands* $G(t)$ are met primarily by the astrocytes, which integrate the required energy consumption over time and space and make a *complementary oxygen demand* $G(t)$ on the

adjacent vasculature. The *hemodynamic processes* $H(t)$ provide the requisite oxygen exchange to replenish the energy depletion in the astrocytes. The fMRI BOLD analysis provides an estimate $Y(t)$ of the ratio of oxygenated to deoxygenated hemoglobin in the blood complement of a given voxel.

MODEL OF THE NEURAL-TO-BOLD TRANSDUCTION CASCADE

The terms in Figure 1 are related by a series of mathematical operations specified in Table 1 (see main text). The three operators are: linear convolution ($\otimes$), a non-linear, zero-memory transducer ($f[\]$) and a multiple linear integrator (Σ). It is important to emphasize that the linear integration across multiple parallel elements in a network corresponds to a non-linear process in the context of single-channel solution. (Linearity is defined as obeying the principle of superposition, such that the response to any combination of inputs is equal to the simple sum of the responses to the individual inputs.) In the non-linear neural-to-BOLD model of Table 1 (see main text), which is considerably elaborated from the conventional BOLD analyses, we make the simplification that each stage consists of the linear convolution of the output signal from the previous stage with a temporal response kernel [which is designated by lower case initial for the name of that stage, i.e., $n(t)$, $m(t)$, $g(t)$, $h(t)$] and the paramagnetic response $p(t)$ that generates the BOLD signal).

Where necessary to model the physiological processes, the input to the linear convolution kernel is first processed with one of the non-linear operators. The last line of the table corresponds to the GLM, in which a function representing the neural metabolic demand evoked by the stimulus presentation is convolved with a MRK. The widespread utilization of the GLM implies that it is an adequate approximation under typical recording conditions, but the focus of this paper will be to emphasize the limitations of this model.

To pursue the analysis of neural responses underlying BOLD signals, we consider the real-data responses to rapid stimulus events in an event-related paradigm (see Methods, main text). The experimentally measured event-related BOLD response.

The model $Y(t)$ is a functional concept that incorporates the full sequence of linear or non-linear neural and metabolic/hemodynamic response properties that follow the stimulus event of whatever form. $Y(t)$ for this paradigm should be distinguished from $h(t)$, the underlying *hemodynamic response function*, or *kernel*. (The hemodynamic response function, or HRF, is the kernel specific to the hemodynamics of blood oxygenation and is based on the assumption that its input is an invariant Dirac pulse.) The empirical response waveform is the averaged BOLD activity synchronized with any repeated stimulus event.

THEORETICAL DEVELOPMENT FOR APPLICATION TO TIME-RESOLVED fMRI

Although the time-resolution of FMRI is low relative to the millisecond time-resolution of single-unit recordings, it must be recognized that many single-unit studies are noise-limited and use response integration up to 500 ms or so, and that many interesting perceptual processes have durations and delays this long and longer. Thus, temporal resolution of fMRI delays in the range below 1 s can deliver important information about the neural signals underlying

such perceptual events, as demonstrated by Menon et al. (1998), in particular. These authors were able to resolve reaction-time delays of the order of 100 ms, and Henson et al. (2002) used the basis function of the first derivative of the MRK (HRF) to determine the sign of the advance or delay of the BOLD response (although the latter approach assumes a unitary MRK in all response regions, which may confound delays with other waveform differences (Bellgowan et al., 2003). Similarly, Sun et al. (2005), working with a frequency domain technique, used the slope of the phase of the fMRI spectrum as an index of relative neural delay between brain areas. This method allowed the mapping of causal influences from a chosen brain areas to the rest of the brain with a typical accuracy of about 500 ms.

The time-domain approach of present analysis allows the extraction of the maximum possible information about the temporal evolution and processing non-linearities of the neural signals underlying particular BOLD activation profiles. Our analysis reveals that much information about the neural response properties are reflected in the BOLD signals, even if the time-resolution is insufficient to reproduce the exact neural signal. Examples of such “transparency” of response properties are particularly clear in the case that the full MRK is monophasic.

This monophasic assumption was tested for rat cortex by both Martindale et al. (2003) and de Zwart et al. (2005), who showed that the empirical dispersion of the BOLD response generated increasing delay with distance from the activation site, but were always well-fit by a monophasic model of the BOLD impulse response. Similarly direct measurements of cerebral blood flow and the concentrations of oxygenated and deoxygenated hemoglobin in the human brain (Hoge et al., 2005) reveal only a monophasic temporal waveform for each of these contributors to the BOLD response. These results are all compatible with the inference of a dominant monophasic positive BOLD response in cat LGN and cortex reviewed above (Thompson et al., 2003, 2004, 2005). This form also seems to be a fair approximation in the case of human fMRI because the canonical response kernel (commonly known as the HRF) provided in the SPM-5 software package, although biphasic, has a negative lobe of less than 10% of the amplitude of the positive lobe. It is therefore only a minor approximation of this standardized kernel to assume that it has no negative lobe, which is the assumption made for the following analysis. (As will be explained below, the residual biphasic component can be readily attributed to neural rebound signals.) Armed with this assumption, we show how several properties of the neural signal are reflected in the recorded BOLD waveform. (For the purpose of this demonstration, we are assuming a linear relationship between the neural response and the BOLD waveform, in order to make its properties clear before introducing the non-linear aspects discussed in the previous section.)

Polarity

The BOLD response polarity will be an accurate reflection of the polarity of the neural response for any monophasic neural response.

Latency

Any delay in the neural response will also be reflected in the consequent BOLD response. Of course, the metabolic processing sequence may introduce additional delays, but neural delays such

as response reaction times or perceptual ambiguity delays should be accurately reflected in the BOLD waveform once the inherent delays of the MRK are taken into account.

Transience

It is an established property of the BOLD response that it is largely sustained for an appropriately sustained neural response (Birn et al., 2001; Boynton et al., 1996; Glover, 1999; Logothetis, 2002, 2003; Shmuel et al., 2006). Hence, transience of the BOLD response for a sustained stimulus implies a transience of the underlying neural response. For example, the onset of a sustained light is known to generate transient neural responses in most cortical neurons responding to it. Such stimulation will generate a transient BOLD response even though the stimulation and photoreceptor response are sustained.

Number of phases

For a monophasic MRK (HRF), the BOLD waveform will have the same number of phases as the input stimulus, if the neural input is balanced for positive and negative lobes. Thus, the fact that the measured BOLD waveform is typically biphasic does not imply that the MRK is necessarily biphasic. The negative lobe may derive from a biphasic neural response to a stimulus rather than to blood dynamics.

Some of these properties of the BOLD response are illustrated in the simulation of **Figure A1**, for which the MRK is assumed to be the gamma function shown in **Figure A1A**, right, as the basis function for the formal analysis. Gamma bases are statistical descriptors of the occurrence of discrete particles (Stacy, 1962; Farwell and Prentice, 1977) that have a simple analog implementation that is a cascade of identical lowpass filters (De Vries and Principe, 1992; Celebi and Principe, 1995; Chen, 2006). In terms of molecular diffusion processes within neurons, the gamma function represents an optimal description of the cooperative process of the arrival of effector molecules at the channels controlling current flow through the cellular membrane (Shao, 1997). We derive the normal equation for the optimum value of the time scale parameter and decouple it from that of the basis weights. Using statistical signal processing tools, we further develop a numerical method for estimating the optimum time scale.

Each left-hand panel shows a different neural waveform with a time-course being a simplified version of a form typical of those seen in neurophysiological recordings. Convolution of each of the waveforms in the left column with a monophasic MRK (HRF) of the form of **Figure A1A** (right panel) generates the predicted BOLD waveforms of the right-hand column of **Figure A1**. The important point made by this simulation is that most typical features of measured BOLD waveforms could arise from convolution of a simple gamma-function MRK with a variety of neural signals modulated within less than 100 ms (**Figures A1A,C,D**). Extended neural signals will generate BOLD waveforms with slower time-courses (**Figure A1B**).

Conversely, under the assumption of a uniform monophasic MRK, the neural signal properties can be inferred from the form of the BOLD response. Thus, if the BOLD response rises to a sustained plateau (**Figure A1B**, right), we may infer that the neural signal was sustained (**Figure A1B**, left); if the BOLD response is biphasic

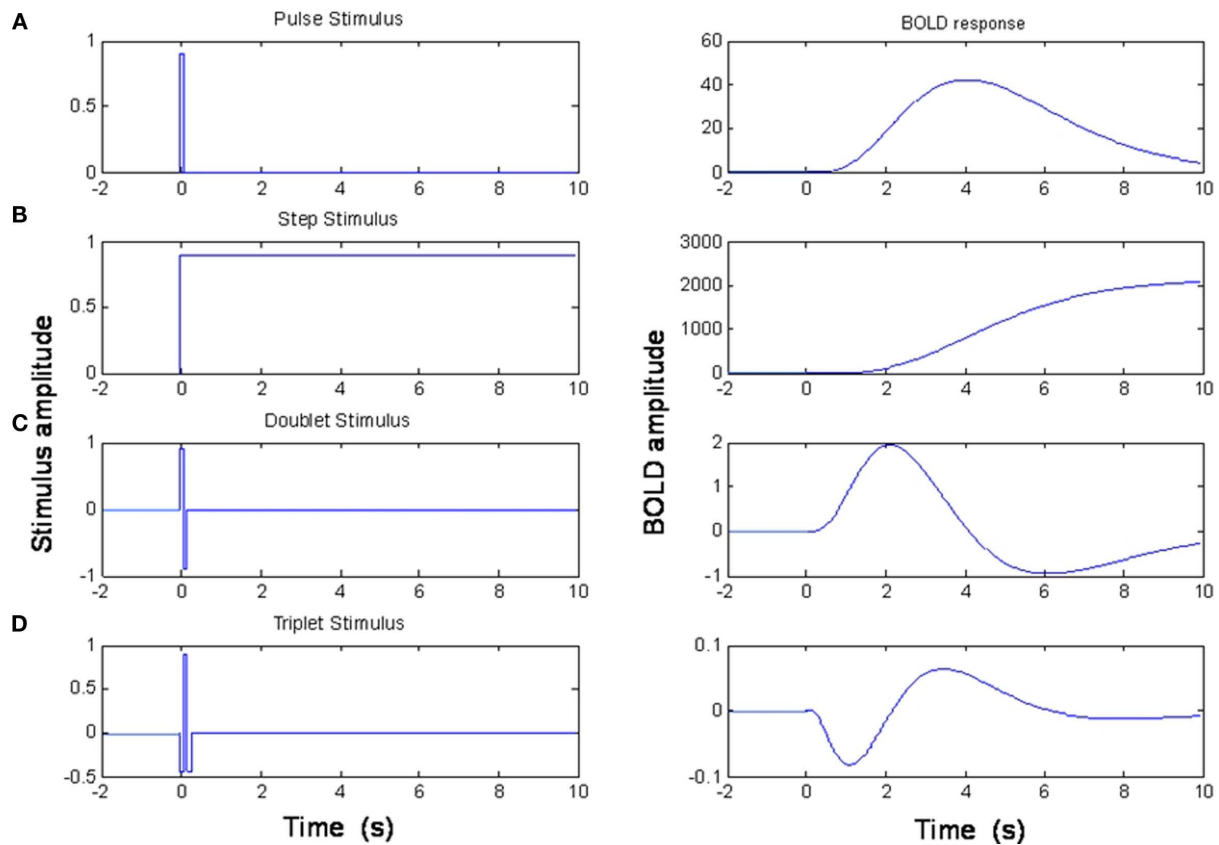


FIGURE A1 | Left panels: (A) neural impulse response, (B) step response, (C) balanced doublet, and (D) balanced triplet response. Right panels present convolution of each of these responses with the MRK shown at right. Note that differences in neural response

characteristics (left panels) at the time scale of 100 ms generate profound changes in the simulated BOLD waveforms (right panels) on a much longer timescale, which in turn are diagnostic of the differences in the neural signals.

(Figure A1C, right), we may infer that the neural signal was biphasic (Figure A1C, left); if the BOLD response is triphasic (Figure A1D, right), we may infer that the neural signal was correspondingly triphasic (Figure A1D, left). Figure A1 thus illustrates that, under appropriate assumptions, the form of the BOLD response can act as a kind of “temporal microscope” for the form of the underlying neural response. Although it cannot resolve the precise temporal parameters (e.g., whether the neural response lobes have durations of 1, 10, or 100 ms), many qualitative aspects of the waveform structure are reflected in the BOLD waveform.

As Figure A1 illustrates, the interpretation is particularly straightforward if the BOLD metabolic/hemodynamic kernel is monophasic. We emphasize that previous suggestions that it is biphasic are contaminated by the likelihood that the underlying neural signal is itself biphasic. Under the logic of the analysis in Figure A1, this property would generate the biphasic form of empirical hemodynamic kernel that is commonly observed, but it seems that previous analyses of BOLD properties have not taken this possibility into account. *In fact*, the biphasic property that is often empirically observed is quite variable over brains and cortical regions (Bellgowan et al., 2003). Based on this analysis, we propose that (i) it may be more plausible to assume that the true metabolic

kernel has a monophasic form, and that (ii) the biphasic form may better be interpreted as a function of the *neural* processing that precedes the metabolic demand, as illustrated in Figure A1, rather than of the metabolic processes.

Specifically, it is well-established that the transmission delay through retina to cortex is about 40 ms in the human brain, and that the monosynaptic transmission delays from one cortical region to another are only about 10 ms in monkey brain (Nowak and Bullier, 1997; Azouz and Gray, 1999), and similarly in human brain. These extra transmission delays may be small in relation to the 70–200 ms peak latencies of the typical neural response in monkey cortex (V1–V4; Hegdé and van Essen, 2004, 2006). The predominant parameter controlling delays of this magnitude is thus the neural time constant τ_m . As will be shown, the neural response parameters for human will be estimated to an accuracy of ~ 0.1 – 0.2 log units, so intracortical transmission delays of a few tens of milliseconds are negligible in this context.

We emphasize that our criterion for the completeness of such a model is that it should be sufficiently well-specified that it characterizes neural responses recorded in monkey cortex, and that its output could be taken as the input to the level of the practical analysis of the neural signals underlying fMRI activation in the human

brain. Although there are numerous analyses of the empirical temporal properties of neural populations, it is surprising that none have taken a mathematical modeling approach to the specification of the temporal response, defining its population parameters in a manner that permits such a neural analysis of the empirical variations. However, without such prior information about the population parameters, such as the range of exponential decays from the transient response peak characteristic of each retinotopic area in monkey cortex, we will not have sufficient power to account for all the parameters of a quantitative model of the response behavior in human cortex.

VALIDATION OF THE NEURAL MODEL WITH THE LOCAL CORTICAL SIGNAL

As is highlighted by the data of Logothetis (2003), there are adaptive effects in the neural response that become evident for long-duration activation by continuous dynamic stimuli (**Figure A2**). The LFP recordings (blue trace) from monkey V1 reveal that the adaptation has a complex time course that can be approximated by two exponentials with time constants of about 1 and 60 s, respectively. These are remarkably prolonged neural processes that are on the time scale of BOLD activation, and will therefore affect the form of the recorded BOLD signal from the same general region of cortex (**Figure A2**, dark gray trace).

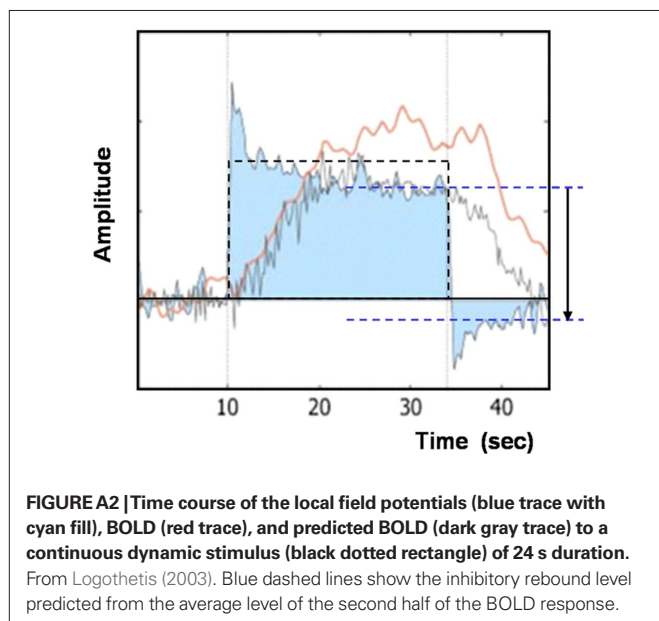


Table A1 | Estimated effect sizes for each parameter.

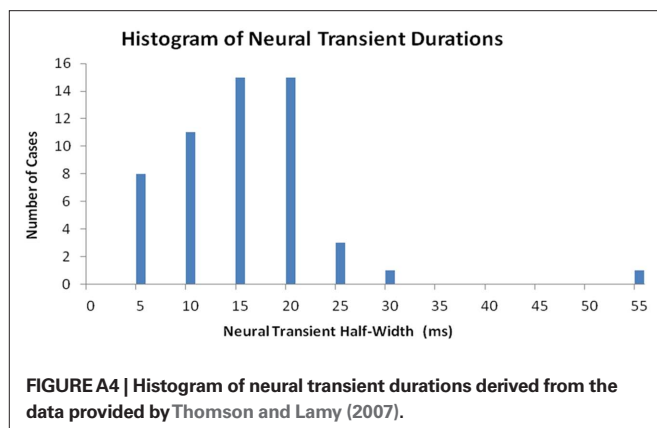
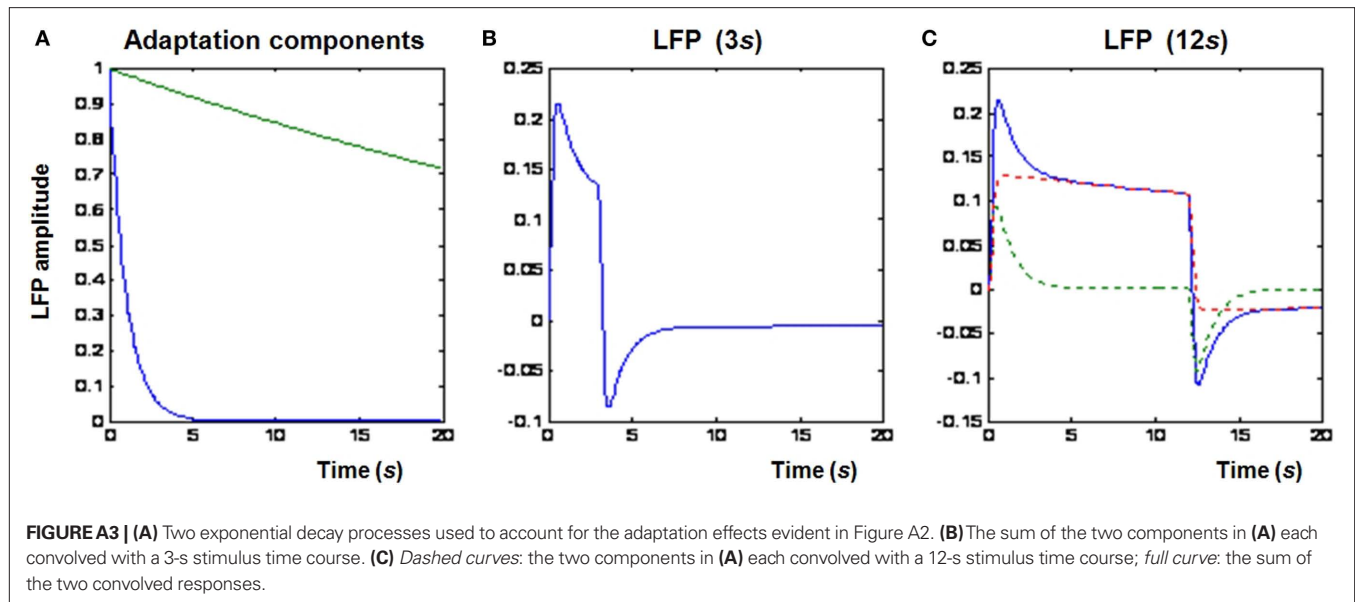
Parameter	Peak latency	S/T ratio	Off/on ratio	MRK width	MRK negative
Estimated variability	+70 ms	+0.30	+0.66	+0.6 s	+0.15
Estimated range	70–200 ms	0–1	–0.5–0.5	4–7 s	0–1
Estimated effect size (Cohen <i>d</i>)	1.86	3.3	1.5	5.0	6.7

The importance of this adaptation effect is emphasized by the fact that the recorded LFP signal does not fully match the BOLD activation predicted (red trace, **Figure A2**) on the basis of GLM of convolution of a hemodynamic response kernel with the stimulus time course, and therefore a more comprehensive model is required, going beyond the standard implementation of the GLM. (This analysis applies when the stimulus is continuously refreshed to support the assumption that the neural signal would be well approximated by the stimulus waveform).

Notice that the negative LFP signal following the stimulus *offset* in **Figure A2** has a transient time course that is an inverted copy of that following the stimulus *onset*. This inversion implies that the rebound effect in the LFP is fundamentally linear. There is also a slow adaptation effect from about 3–24 s, with a corresponding recovery following the offset transient. This adaptation effect is a *subtractive* inhibition rather than a solely multiplicative form of fatigue (which would have no negative rebound). The predicted offset for a linear system from the adapted level at 24 s is shown by the blue dashed lines, which give an accurate prediction of the offset level following the initial offset transient. If there were a multiplicative gain control, the signal change following offset would be substantially less than that at onset, whereas the two amplitudes are quite similar.

The mechanism of this subtractive inhibition appears to be the tonic intracellular hyperpolarization induced by pattern adaptation, as described by Carandini and Ferster (1997, 2000). However, the stimulus driving the response in **Figure A2** was a *dynamic* contrast modulation, implying that the sustained LFP signal derives from a full-wave rectified transform of the intracellular potential. This analysis leads to a model of the LFP as the integrated response of instantaneous signal of the *rectified* intracellular potential from the nearby neurons, convolved with an exponential decay function. However, the data in **Figure A2** clearly show a short decay to a stable response level followed by a slow decay of about 10% over the 30 s of the full stimulus epoch. This dual decay characteristic can be fit by a *parallel-process decay model*, in which the two decays come from separate parallel processes summed into the measured signal.

An implementation of the parallel-process model that we propose on this basis is shown in **Figure A3**, using the parameters for the qualitative fit to the data of **Figure A2**. It incorporates two decaying exponential components with time constants of 1 and 60 s (**Figure A3A**). These components are each convolved with a neural signal derived from the convolution of the rectangular (box-car) form of the continuous stimuli of 3 and 12 s duration (cf. Logothetis, 2003). The latter stimulus was also the one used in **Figure A2**. This parallel-process model (full curve in **Figure A3C**) captures the qualitative features of the LFP data (**Figure A2**) with the sum of the two component response (dashed curves in **Figure A3C**). These parallel components represent the average responses of two separate neural populations contributing to the recorded LFP. It is problematic to obtain the integrated response, i.e., the combination of the two component slopes of **Figure A3C**, with any kind of serial model, because this would imply a convolution of the two exponentials that would necessarily result in a function dominated by the faster process rather than allowing both processes full expression.



FMRI METHODS

Functional MRI data were acquired with a 3-T magnetic resonance (MR) scanner (Siemens Trio) using a single-shot gradient echo-planar imaging (EPI) of 34 slices acquired inferior to superior in interleaved order with no gap. TR = 2000 ms, TE = 27 ms, flip angle = 90°, in-plane FOV = 220 mm × 220 mm with resolution of 96 × 96 yielding 2 mm × 2 mm × 2 mm voxels throughout the occipital lobe. The phase encoding direction was right-to-left. The scan was followed by a 2-D spin echo T1 scan with the same in-plane prescription. (The subject was also scanned with a high-resolution T1 with a 3-D MP-RAGE sequence with TR = 2300 ms, TE = 2.96 ms, flip angle = 9° in 208 sagittal slices with 256 × 256 mm FOV and 320 × 320 resolution yielding 0.8 mm × 0.8 mm × 0.8 mm voxels.)

The functional activations were then processed for slice-time correction, motion correction, and trend removal to minimize the low-frequency drift in the BOLD signal. Activations were analyzed only for voxels lying within the cortical gray matter, as defined by an anatomical segmentation algorithm (mrGray), which ensures localization of the signal close to the activated neurons and greatly reduces the blood drain artifacts. The 228 s time series for each

cortical voxel were averaged across 12 scans at the fMRI sampling rate of 2 s. Temporal jitter over a range of ±3 s in the presentation times was implemented on a 1-s time base so that the 2-s samples of BOLD activation could be interleaved to a 1-s sampling rate.

PERTURBATION ANALYSIS OF THE MODEL FITS AND ESTIMATION OF EFFECT SIZES

This qualitative description of the typical response parameters may be further refined with a perturbation analysis to reveal the sensitivity of each parameter (Bevington and Robinson, 2003). Once the optimum fit is achieved, each parameter is perturbed to determine its sensitivity for degrading the fit, which is a form of power analysis. The procedure for this determination was to determine the sensitivity for a given parameter the model was refit with that parameter set at a range of values, with all the other parameters varying at each value of the given parameter. Thus, the specified sensitivity ranges take into account any potential trade-offs among parameters.

If small perturbations generate strong degradation of the fit, the neural response parameter is well-constrained by the BOLD dataset and its value has been determined with high confidence. The criterion for the perturbation error defining the parameter sensitivity is the perturbation level at which the %² fit becomes significantly worse at $p < 0.05$. Perturbation analysis of the parameters for the fits in **Figure 4** of the main text shows that the transient width (neural integration time) estimate is accurate to about ±70 ms, and the sustained/transient peak ratio to about ±30%. These are remarkably tight constraints, as are those of the BOLD MRK parameters of net integration time (±0.6 s), the negative BOLD response ratio (MRK neg) (±15%), and the weight on the neural off-response rectification ratio to about ±66%.

These computed levels of variability may be converted to estimates of effect size based on the expected range of variation of each parameter among cortical areas (from extensive published work in monkey cortex). The peak latency (x) is derived according to the

analysis of **Figure 3** in main text. These estimated effect sizes (from Cohen, 1992), tabulated in **Table A1**, are impressively large for most of the parameters, since any effect size greater than 1 means that a parameter value can be determined to within its expected range of physiological variation.

It should be emphasized that no previous study on any kind of neural signal estimation has previously conceptualized how to achieve this level of temporal resolution from fMRI measurements. The NDFO approach of **Figure 5** of the main text has essentially unlimited ability to resolve the neural temporal characteristics of the order of a few milliseconds from the BOLD temporal duration series. All that is needed is the ability to present stimuli at durations spanning those of the neural response, and appropriate levels of signal/noise ratio of the BOLD (e.g., SNR > 10:1).

ANALYSIS OF NEURAL TIME CONSTANTS

To assess the plausibility of the peak latency estimate in **Table 1** in main text, we provide an analysis of the characteristics of neuronal response in more than 50 studies of intracellular and whole-cell current recordings in cat, rat, and a few monkey species tabulated in a recent paper by Thomson and Lamy (2007). Despite the species differences, the results indicate a relatively tight constraint on the neuronal response dynamics, which we have summarized in a frequency histogram of the durations of the neuronal transients (**Figure A4**). This histogram is derived from half-widths of the (intracellular) excitatory or inhibitory post-synaptic response transients in the primary sensory cortical regions. From this summary we infer that that typical transient half-width is of the order of 18 ms, corresponding to standard deviation of ~20 ms at half height.